



<https://www.helicojournal.org>

Received August 5, 2023
Revised September 3, 2023
Accepted September 5, 2023

Corresponding author
Soo-Jeong Cho, MD, PhD
Division of Gastroenterology,
Department of Internal Medicine and
Liver Research Institute,
Seoul National University Hospital,
Seoul National University
College of Medicine,
101 Daehak-ro, Jongno-gu,
Seoul 03080, Korea
E-mail: crystal5@snu.ac.kr

Availability of Data and Material
The datasets generated or analyzed during the study are available from the corresponding author on reasonable request.

Conflicts of Interest
Soo-Jeong Cho, a contributing editor of *The Korean Journal of Helicobacter and Upper Gastrointestinal Research*, was not involved in the editorial evaluation or decision to publish this article. All remaining authors have declared no conflicts of interest.

Funding Statement
None

Acknowledgements
None

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<https://creativecommons.org/licenses/by-nc/4.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Types of 23S Ribosomal RNA Point Mutations Affecting *Helicobacter pylori* Eradication Rates in Clarithromycin-Based Triple Therapy

Gihong Park, Bokyoung Kim, Hyunsoo Chung,
Sang Gyun Kim, and Soo-Jeong Cho

Department of Internal Medicine and Liver Research Institute, Seoul National University Hospital, Seoul National University College of Medicine, Seoul, Korea

Objectives: The A2142G and A2143G mutations in the 23S ribosomal ribonucleic acid (rRNA) of *Helicobacter pylori* are the most common mutations associated with clarithromycin resistance. This study aimed to determine the differences in *H. pylori* eradication rates in patients infected with bacteria carrying the A2142G and A2143G mutations who were treated with clarithromycin-based triple therapy. **Methods:** Data from a previous randomized controlled trial were analyzed retrospectively. Eradication rates were compared based on the presence of *H. pylori* carrying the A2142G and A2143G mutations. A meta-analysis was also conducted of relevant studies containing data regarding patients who received clarithromycin-based therapy due to infections with *H. pylori* harboring 23S rRNA mutations. **Results:** No significant difference was observed in *H. pylori* eradication rates between patients infected with wild-type bacteria (95.7% [44/46]) compared with those infected with bacteria carrying the A2142G mutation (100.0% [3/3]; $p>0.9$). However, the eradication rate was significantly lower for patients infected with bacteria carrying the A2143G mutation (16.7% [1/6]; $p<0.001$) than for those infected with wild-type bacteria or bacteria with the A2142G mutation (100.0% [3/3]; $p=0.048$). In the meta-analysis, the between-group comparisons yielded similar results. Although patients infected with bacteria having the A2142G mutation exhibited no significant risk difference (RD) for eradication compared with those infected with wild-type bacteria (RD=-0.05 [-0.18 to 0.08]; $I^2=0\%$; $p=0.42$), those infected with bacteria having the A2143G mutation demonstrated a lower *H. pylori* eradication rate compared with patients infected with either wild-type (RD=0.72 [0.64-0.80]; $I^2=0\%$; $p<0.001$) or A2143G mutant bacteria (RD=0.76 [0.61-0.91]; $I^2=0\%$; $p<0.001$). **Conclusions:** The A2143G mutation may play a more significant role in clarithromycin triple therapy *H. pylori* eradication failure than does the A2142G mutation. Additionally, *H. pylori* strains with the A2142G mutation can be treated effectively with clarithromycin-based triple therapy.

Keywords *Helicobacter pylori*; Clarithromycin; Polymerase chain reaction; Drug resistance, microbial.

INTRODUCTION

Helicobacter pylori infection is the major cause of gastrointestinal diseases such as chronic gastritis, peptic ulcer disease,

gastric mucosa-associated lymphoid tissue lymphoma, and gastric adenocarcinoma.¹⁻³ Eradication of *H. pylori* prevents long-term complications and recurrence of gastrointestinal diseases caused by the infection.^{1,3-5} Standard triple therapy (stan-

standard-dose proton pump inhibitor [PPI], amoxicillin [1 g], and clarithromycin [500 mg] twice daily for 7 days) has been widely prescribed and recommended as a first-line treatment in Korea since 1998.⁶ However, the rate of clarithromycin resistance has increased continuously. A nationwide resistance study, conducted in 2018, reported a resistance rate of 17.8%–31%.⁷ In the Maastricht V/Florence consensus report, therapies other than the standard triple therapy are recommended in areas with high (>15%) clarithromycin resistance rates.⁸ According to the 2021 Korean guidelines, a clarithromycin resistance test determined using polymerase chain reaction (PCR) or sequencing is recommended when 7-day standard triple therapy is considered as a first-line treatment.⁶

Point mutations in the peptidyl transferase region encoded in domain V of the *H. pylori* 23S ribosomal ribonucleic acid (rRNA) are crucial for clarithromycin resistance.⁹ Several studies have demonstrated that among the point mutations known to cause clarithromycin resistance, the A2142G and A2143G mutations are the most common.^{9,10} Among the available molecular tests, dual-priming oligonucleotide multiplex PCR (DPO-PCR) has been commercialized and used in clinical practice and various clinical studies to detect A2142G and A2143G mutations. In multiple studies, a concordance rate of over 90% has been observed between DPO-PCR and culture-based susceptibility testing.^{11,12}

A randomized controlled trial (RCT) was conducted at Seoul National University Hospital from January 2019 to June 2019 to evaluate the efficacy of clarithromycin resistance-guided tailored therapy using the DPO-PCR test.¹³ The present study aimed to retrospectively compare the effects associated with the A2142G and A2143G mutations on *H. pylori* eradication rates following clarithromycin-based triple therapy using a subgroup analysis of the previous RCT. A systematic literature review was also conducted to analyze relevant studies related to this topic.

METHODS

Trial design and patient selection for subgroup analysis

Among the patients in the previous RCT, those who were treated with clarithromycin-based triple therapy (esomeprazole [40 mg], amoxicillin [1 g], and clarithromycin [500 mg] twice daily for 10 days), regardless of the mutational status of the infecting *H. pylori* as determined by DPO-PCR, were selected for statistical analysis. As this study aimed to compare the contributory importance of each point mutation on the *H. pylori* eradication rate for clarithromycin-based triple therapy, the analysis was performed using the per-protocol method. All patients selected for subgroup analysis underwent a urea

breath test (UBT) to confirm eradication. The eradication status was determined using ¹³C-UBT conducted at least 4 weeks after treatment completion. Patients were required to discontinue PPI or histamine-2 receptor blocker use for at least 2 weeks before the test.

Determination of clarithromycin resistance

Gastric biopsy specimens were obtained from the stomach antrum and body of each patient. Moreover, DPO-PCR was performed on the biopsy specimens using a commercial kit (Seeplex ClaR-H. *pylori* ACE Detection; Seegene Institute of Life Science, Seoul, Korea). The DPO-PCR results were interpreted as follows: wild-type bacteria were indicated by a single 621-bp deoxyribonucleic acid (DNA) product, bacteria harboring the A2142G mutation by a DNA band at 475 bp, and bacteria with the A2143G mutation by a DNA band at 194 bp. Detailed information about the PCR analysis is described in the previous study report.¹³

Method for systematic search of relevant studies and meta-analysis

To find relevant studies, a systematic search was conducted on PubMed using the following keywords: “A2142G,” “A2143G,” “*Helicobacter pylori*,” and “Eradication.” Following a search using the aforementioned keywords, each article was reviewed to select studies containing data regarding patients infected with *H. pylori* strains carrying the A2142G or A2143G mutations and treated with clarithromycin-based triple therapy. Studies focusing solely on in vitro resistance profile tests, without eradication treatment, were excluded. Using this method, we aimed to identify studies specifically focusing on assessing the efficacy of clarithromycin-based triple therapy in patients infected with *H. pylori* strains carrying the A2142G and A2143G mutations. A meta-analysis was conducted to calculate pooled estimates of the eradication rates and to perform between-group comparisons for each mutation.

Statistical analysis

Fisher’s exact test and the chi-square test were used to compare categorical variables. The Wilcoxon rank-sum test was conducted to compare continuous variables exhibiting a non-normal distribution. For the meta-analysis, a random-effects model was considered most appropriate because of the differing study designs, diverse study objectives, and variations in patient characteristics across the included studies. All statistical analyses were performed using R software, version 4.3.1 for Windows (R Foundation for Statistical Computing, Vienna, Austria). The meta-analysis was performed using the “meta” package of the R software. Moreover, *p*-values less than 0.05

were considered statistically significant.

Ethics statement

This study was approved by the Institutional Review Board of Seoul National University Hospital, Seoul, Korea (2307-070-1448). The requirement for written consent was waived owing to the retrospective nature of this study which used data obtained from a previous RCT and a literature review of other relevant studies. This study was conducted following the principles of the Declaration of Helsinki.

RESULTS

Retrospective analysis of a previous RCT

Among the 72 patients enrolled in the previous RCT,¹³ nine were identified as being infected with *H. pylori* carrying either the A2142G or A2143G mutation; three were infected with *H. pylori* carrying the A2142G mutation, five with bacteria carrying the A2143G mutation, and one with bacteria carrying both mutations (classified into the A2143G mutation group, consistent with the study purpose and the analysis of other relevant studies^{14–18}). These patients subsequently received clarithromycin-based triple therapy. The baseline characteristics of the patients included in the subgroup analyses are displayed in Table 1. No significant differences in the baseline characteristics were observed between the groups.

Additionally, no statistically significant differences were observed in the *H. pylori* eradication rates between patients affected by the wild-type bacteria and those infected with bacteria carrying A2142G mutation (95.7% [44/46] vs. 100.0% [3/3]; $p>0.9$). In contrast, patients infected with bacteria carrying the A2143G mutation exhibited a significantly lower eradication rate (16.7% [1/6]) than did those infected with wild-type bacteria (95.7% [44/46]; $p<0.001$) or those infected with bacteria carrying the A2142G mutation (100.0% [3/3]; $p=0.048$). A graphical representation of the results is illustrated in Fig. 1.

Literature search and meta-analysis of other related studies

A literature search was conducted using the aforementioned keywords, yielding a total of 77 articles. In most of these studies, patients infected with *H. pylori* strains carrying point mutations were not treated with clarithromycin-based therapy because of antibiotic resistance. After excluding studies that did not meet the prespecified criteria, five articles were identified. In three articles, the eradication rates were directly compared between patients infected with the A2142G and A2143G mutants. In the other two articles, although the primary goal was not to directly compare *H. pylori* eradication rates between groups of patients infected with the two mutant strains, subgroup analyses of the patient data were available.

In a study conducted by Jung et al.,¹⁹ 145 patients were infected with wild-type *H. pylori*, 28 with bacteria carrying the A2143G mutation, and three with bacteria carrying the A2142G mutation. All patients were treated with standard triple therapy. The eradication rates did not differ between the patients infected with wild-type or A2142G mutant *H. pylori* (93.1% [135/145] vs. 100.0% [3/3], respectively; $p>0.9$). In contrast, the eradication rate associated with patients infected with A2143G

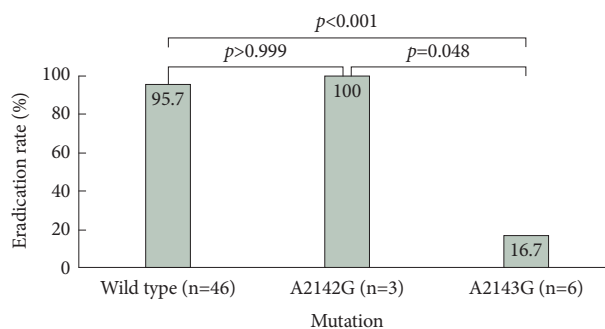


Fig. 1. Mutation and eradication rate. Statistical comparison of the *Helicobacter pylori* eradication rates among patients infected with the wild-type, A2142G mutant, and A2143G mutant bacteria. A $p<0.05$ is considered statistically significant. Fisher's exact test is used for analysis.

Table 1. Baseline characteristics of patients included in the retrospective analysis

Characteristics	Wild type (n=46)	A2142G mutation (n=3)	A2143G mutation (n=6)	p-value*
Sex				0.613
Male	24 (52.2)	2 (66.7)	5 (83.3)	
Female	22 (47.8)	1 (33.3)	1 (16.7)	
Age (yr)	65.0 (60.5–73.5)	71.0 (59.5–74.0)	64.0 (62.0–77.0)	0.834
Hypertension	26 (56.5)	2 (66.7)	3 (50.0)	>0.999
Diabetes	10 (21.7)	1 (33.3)	2 (33.3)	0.412
Smoking	13 (28.3)	1 (33.3)	3 (50.0)	0.395
Drinking	16 (34.8)	3 (100.0)	2 (33.3)	0.061

Data are presented as n (%) or median (IQR).

*Fisher's exact test for categorical variables and Kruskal-Wallis rank sum test for a continuous variable with non-normal distribution. IQR, inter quartile range.

mutants was significantly lower (28.5% [8/28]) than those of patients infected with A2142G mutants (100.0% [3/3]; $p=0.036$) or wild-type *H. pylori* (93.1% [135/145]; $p<0.001$).

In a study conducted by De Francesco et al.,¹⁵ 75 patients were assigned a 7-day standard triple therapy, including 59 patients infected with wild-type bacteria, 10 with bacteria having the A2143G mutation, and six with bacteria carrying the A2142G mutation. The *H. pylori* eradication rates were not different between patients infected with wild-type bacteria and those infected with A2142G mutants (86.4% [51/59] vs. 83.3% [5/6], respectively; $p>0.9$). In contrast, the eradication rate associated with patients infected with A2143G mutants (20.0% [2/10]) was significantly lower than those of individuals infected with either A2142G mutants (83.3% [5/6]; $p=0.035$) or wild-type bacteria (86.4% [51/59]; $p<0.001$).

Francavilla et al.¹⁶ conducted a similar study using the same methodology as De Francesco et al.¹⁵ In their study, 73 patients were assigned to the standard triple therapy group, including 58 infected with wild-type *H. pylori*, 13 with A2143G mutants, and two with A2142G mutants. The bacterial eradication rates were similar for patients infected with wild-type bacteria (82.8% [48/58]) and those infected with A2142G mutants (100.0% [2/2]; $p>0.9$). However, the eradication rate for patients infected with A2143G mutants (0.0% [0/13]) was significantly lower than the rate for patients infected with A2142G mutants (100.0% [2/2]; $p=0.010$) or wild-type *H. pylori* (82.8% [48/58]; $p<0.001$).

In a study by Kim et al.,¹⁷ 110 patients infected with *H. pylori* were treated with clarithromycin-based triple therapy. Information regarding the genotypic resistance profiles was obtained using DPO-PCR. Seven patients were infected with *H. pylori* carrying the A2142G mutation and 16 were infected with *H. pylori* carrying the A2143G mutation. No differences in bacterial eradication rates were observed between the patients infected with wild-type bacteria (84.7% [50/59]) and those infected with A2142G mutants (85.7% [6/7]; $p>0.9$). In contrast, the eradication rate for patients infected with A2143G mutants was significantly lower (12.5% [2/16]) than that for patients infected with either A2142G mutants (85.7% [6/7]; $p=0.010$) or wild-type bacteria (84.7% [50/59]; $p<0.001$).

In another study conducted by Kim et al.,¹⁸ 464 patients were retrospectively analyzed; 175 were infected with *H. pylori* harboring 23S rRNA mutations. Although the patients were assigned for treatment with bismuth-quadruple therapy based on their genotypic resistance status, 37 underwent clarithromycin-based triple therapy, despite the mutational status of the infecting bacteria; the therapy was prescribed by non-gastrointestinal specialists unfamiliar with the 23S rRNA mutation test. A subgroup analysis of those patients displayed no significant difference in the *H. pylori* eradication rates between the

patients infected with wild-type bacteria (89.8% [229/255]) and those infected with A2142G mutants (100.0% [4/4]; $p>0.9$); a lower eradication rate was observed for patients in the A2143G *H. pylori* group (25.8% [8/31]) than in either the A2142G mutant ($p=0.009$) or wild-type *H. pylori* infection groups ($p<0.001$). A summary of the results of each study and the specifics of the eradication regimens used are provided in Table 2.

A meta-analysis of the six studies was performed using a random-effects model. Pooled analyses of the *H. pylori* eradication rates for each group yielded the following: 90.1% (95% confidence interval [CI], 86.6%–93.6%; $I^2=46\%$) for the wild-type bacterial infection group, 93.9% (95% CI, 81.5%–100%; $I^2=0\%$) for the A2142G mutant infection group, and 16.0% (95% CI, 5.3%–26.7%; $I^2=62\%$) for the A2143G mutant infection group. Between-group comparisons using the same statistical model yielded similar results. Although the patients infected with A2142G mutants exhibited no significant difference in their *H. pylori* eradication rate compared with those infected with wild-type bacteria (risk difference [RD]=−0.05 [95% CI, −0.18 to 0.08]; $I^2=0\%$; $p=0.42$), patients infected with A2143G mutants demonstrated a lower eradication rate compared with either those infected with wild-type bacteria (RD=0.72 [95% CI, 0.64–0.80]; $I^2=0\%$; $p<0.001$) or A2143G mutants (RD=0.76 [95% CI, 0.61–0.91]; $I^2=0\%$; $p<0.001$). The pooled analysis of the eradication rates and the between-group comparisons are displayed in Figs. 2 and 3, respectively.

DISCUSSION

This study demonstrated that, compared with the wild type, the A2143G mutation played a more crucial role in *H. pylori* eradication treatment failure than did the A2142G mutation. This result was consistent with other studies, including three studies conducted in Korea.^{17–19} Among the various 23S rRNA point mutations identified from in vitro studies, the A2142G, A2142C, and A2143G mutations are commonly associated with high-level clarithromycin resistance. The T2182C, A2115G, G2141A, A2144G, and T2289C mutations have also been reported; however, their contribution to clarithromycin resistance is considered low, and their significance in the development of resistance is not fully understood.^{20,21} Given the high prevalence of the A2142G, A2143G, and A2142C mutations and the high resistance rates associated with them, most of the commercialized kits designed to detect clarithromycin resistance related to 23S rRNA mutations involve tests targeting these mutations (Table 3).^{9,22,23}

Although A2142G, A2143G, and A2142C are high-level resistance-related mutations, clinical studies that have specifically investigated the eradication rates of clarithromycin-based

Table 2. *H. pylori* eradication rate of clarithromycin-based triple therapy according to 23S rRNA point mutations

Studies	Eradication regimen	Wild type	Mutation overall	A2142G mutation	A2143G mutation or double
This study	Esomeprazole 40 mg Amoxicillin 1 g Clarithromycin 500 mg Twice daily for 10 days	44/46 (95.6)*†	4/9 (44.4)	3/3 (100.0)*‡	1/6 (16.7)†‡
Kim et al. ¹⁸ (2021)	(Lansoprazole 30 mg or Pantoprazole 40 mg or Rabeprazole 20 mg) Amoxicillin 1 g Clarithromycin 500 mg Twice daily for 7 days	229/255 (89.8)*†	12/35 (34.3)	4/4 (100.0)*‡	8/31 (25.8)†‡
Jung et al. ¹⁹ (2014)	Standard dose PPI Amoxicillin 1 g Clarithromycin 500 mg Twice daily for 7 days	135/145 (93.1)*†	11/31 (35.5)	3/3 (100.0)*‡	8/28 (28.5)†‡
Kim et al. ¹⁷ (2013)	Lansoprazole 30 mg Amoxicillin 1g Clarithromycin 500 mg Twice daily for 7 days	50/59 (84.7)*†	8/23 (34.8)	6/7 (85.7)*‡	2/16 (12.5)†‡
Francavilla et al. ¹⁶ (2010)	Omeprazole (1 mg/kg/day) Amoxicillin (50 mg/kg/day) Clarithromycin (15 mg/kg/day) for 7 days	48/58 (82.8)*†	2/15 (13.3)	2/2 (100.0)*‡	0/13 (0.0)†‡
De Francesco et al. ¹⁵ (2006)	Rabeprazole 20 mg Amoxicillin 1g Clarithromycin 500 mg Twice daily for 7 days	51/59 (86.4)*†	7/16 (43.8)	5/6 (83.3)*‡	2/10 (20.0)†‡
Overall		557/622 (89.5)	44/129 (34.1)	23/25 (92.0)	21/104 (20.2)

Data are presented as n/total n (%). Fisher's exact test was used due to low expected values.

*No statistically significant difference in the eradication rate between the wild-type group and the A2142G group ($p>0.9$); †The A2143G group demonstrated a significantly lower eradication rate compared to the wild-type group ($p<0.05$); ‡The A2143G group demonstrated a significantly lower eradication rate compared to the A2142G group ($p<0.05$).

PPI, proton pump inhibitor.

triple therapy involving these mutations in real-world settings are limited. Most studies have focused on the presence or absence of these mutations as predictive markers for clarithromycin resistance rather than directly comparing eradication rates among patients infected with *H. pylori* harboring the different mutations. In the present study, a direct statistical comparison of the clinical outcomes between mutations was performed. Considering the limitations of the small sample sizes, a review of other related studies was also conducted and analyzed. By including a comprehensive analysis of related studies, the A2143G mutation can be inferred to be a significant mutation contributing to eradication failure. The accumulated evidence from multiple studies strengthens this conclusion, despite the limitations of each study.

The mechanisms explaining the discrepancy between real-

world treatment outcomes and in vitro studies regarding clarithromycin resistance are yet to be identified. Point mutations in domain V of the 23S rRNA can cause structural changes that hinder clarithromycin binding (the primary resistance mechanism).²⁰ However, many other mechanisms also contribute to the development of resistance. One study demonstrated that different mutations in genes outside the 23S rRNA, such as *rpl22* and *infB*, can mitigate clarithromycin resistance, even in the presence of 23S rRNA mutations.²⁴ Several studies have also revealed that the level of clarithromycin resistance in *H. pylori* strains carrying 23S rRNA mutation varies depending on the expression of efflux pumps.^{25,26} Additionally, a recent study has demonstrated that bacterial biofilms are essential for the development of antibiotic resistance.²⁷ A study that utilized a computerized three-dimensional molecular struc-

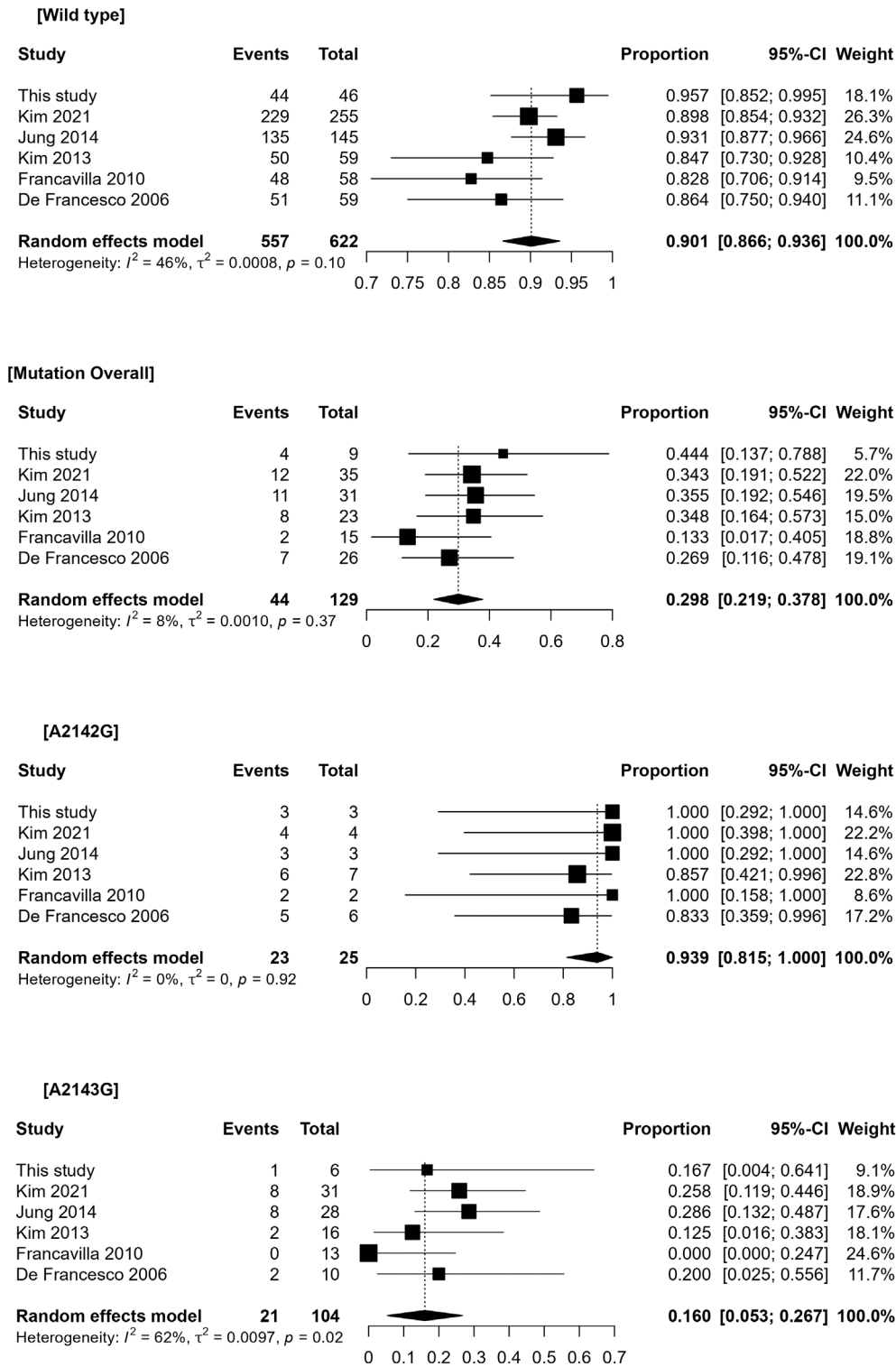


Fig. 2. Pooled analysis of eradication rates.¹⁵⁻¹⁹ CI, confidence interval.

ture prediction model demonstrated that 23S rRNA mutations hinder the binding of clarithromycin and induce secondary structural changes in other areas, including the nascent peptide exit tunnel, which is a crucial component of protein syn-

thesis.²⁸ In an in vitro study, PPIs and amoxicillin have exerted synergistic effects with clarithromycin.²⁹ In addition to the possible causes mentioned above, various factors, including intragastric pH and several other host factors, are presumed to

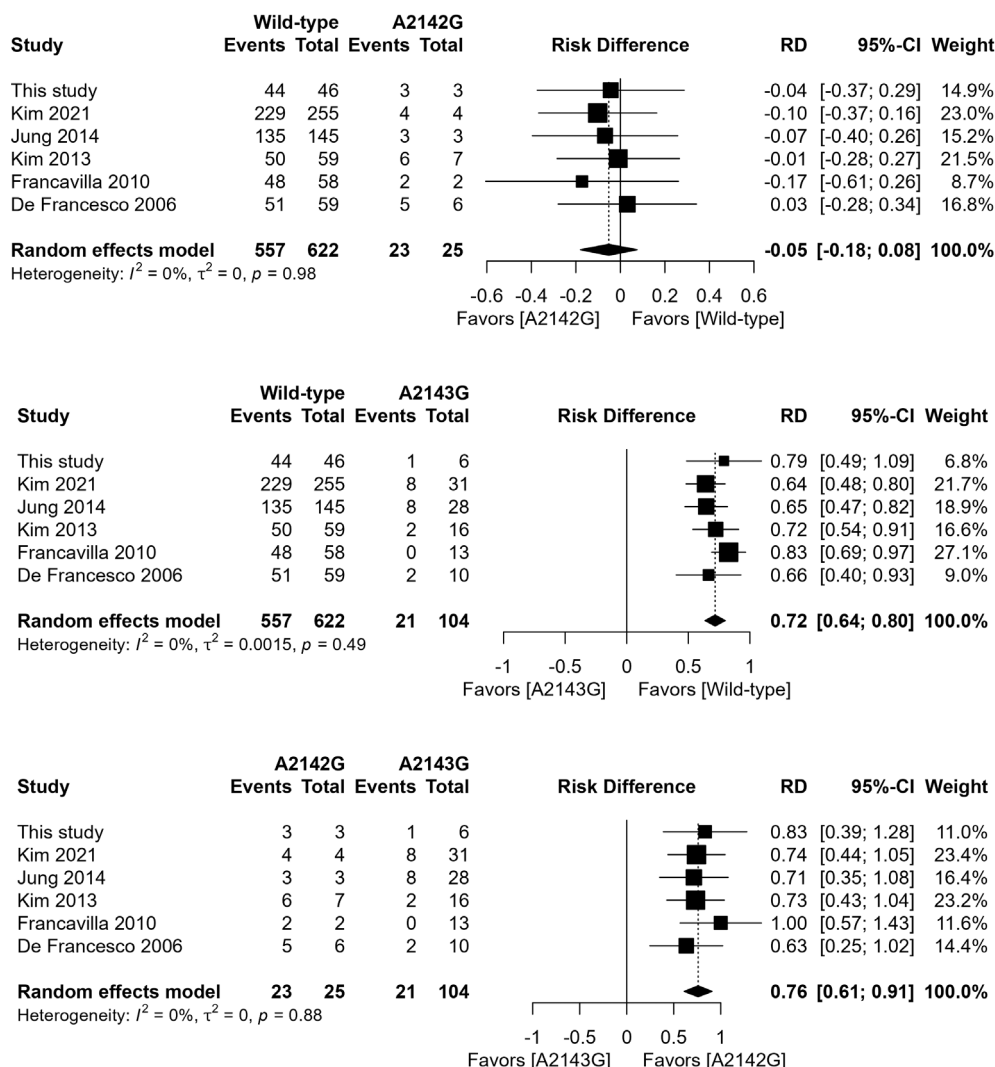


Fig. 3. Between-group comparisons.¹⁵⁻¹⁹ CI, confidence interval.

contribute to the differences in eradication rates. Future studies are needed to identify the reasons for the discrepancies between the findings of in vitro studies and the results of this analysis, including real-world clinical outcomes.

Most current guidelines recommend using non-clarithromycin-based therapies in regions where clarithromycin resistance is reported to be greater than 15% or when resistance is confirmed during testing.^{6,8} However, this subgroup analysis and review of related studies suggested that standard clarithromycin triple therapy for 7 days remains a viable treatment option for patients with the A2142G mutation. Treatment regimens such as bismuth/metronidazole-based therapy have displayed excellent clinical outcomes for the treatment of clarithromycin-resistant strains.^{30,31} However, a recent meta-analysis conducted in Korea exhibited a higher rate of adverse events and drug intolerance associated with quadruple therapy than with clarithromycin-based triple therapy.⁶ Several studies have also

reported increasing resistance rates to metronidazole and tetracycline, which could lower the eradication rate in the future.^{32,33} Considering these factors, diversifying and preserving treatment options for *H. pylori* strains carrying the A2142G mutation may have clinical benefits.

Our study has certain limitations. First, the study was conducted retrospectively, which may have introduced a selection bias. Additionally, the analysis was performed using a per-protocol approach, including patients who were followed up and tested to confirm eradication, which may have overestimated the results. Second, although the differences in eradication rates were statistically significant, the sample sizes were small. However, statistical analyses were conducted on related studies, and the pooled results displayed findings similar to those of our analysis. When drawing conclusions from the meta-analysis, caution is necessary owing to the limited number of available studies, small sample sizes within each study, and the retro-

Table 3. Commercialized products to detect 23S rRNA mutations

	Allplex	Seeplex	ClariRes	GENECUBE	HelicoDR	MutaREAL
Manufacturer (country)	Seegene (Korea)	Seegene (Korea)	Ingenetix (Austria)	TOYOBO (Japan)	Hain Life Science (Germany)	Inmundiagnostik (Germany)
Assay technique	Real-time PCR	Dual priming oligonucleotide PCR	Real-time PCR	Real-time PCR	DNA strip genotype test combining PCR and hybridization	Real-time PCR
Target mutation	23S rRNA A2142G A2143G	23S rRNA A2142G A2143G	23S rRNA A2142G A2143G A2142C	23S rRNA A2142G A2143G	23S rRNA A2142G A2143G A2142C	23S rRNA A2142G A2143G A2142C

spective design of all the included studies. Further research with larger patient cohorts and more robust study designs is warranted to better understand the effect of these mutations on treatment outcomes in clinical practice. These studies can provide valuable insights into the associations between specific 23S rRNA mutations and treatment success, ultimately leading to optimized treatment strategies for *H. pylori* eradication.

Conclusion

The eradication success rate was significantly lower in patients infected with *H. pylori* strains carrying the A2143G mutation than in those infected with wild-type bacteria or bacteria carrying the A2142G mutation. The eradication rate for strains with the A2142G mutation was not significantly different from that of the wild-type strain. Several other studies have displayed similar results, indicating the possibility of treating *H. pylori* strains carrying the A2142G mutation with standard 7-day clarithromycin triple therapy. These results also indicate the potential value of developing commercial kits that include crucial resistance-related point mutations other than the A2142G mutation in 23S rRNA.

Authors' Contribution

Conceptualization: Soo-Jeong Cho. Data curation: Bokyung Kim, Hyunsoo Chung, Sang Gyun Kim, Soo-Jeong Cho. Formal analysis: Gihong Park. Investigation: Gihong Park. Methodology: Sang Gyun Kim, Soo-Jeong Cho. Project administration: Sang Gyun Kim, Soo-Jeong Cho. Resources: Soo-Jeong Cho. Software: Gihong Park. Supervision: Soo-Jeong Cho. Validation: Soo-Jeong Cho. Visualization: Gihong Park. Writing—original draft: Gihong Park. Writing—review & editing: all authors. Approval of final manuscript: all authors.

ORCID iDs

Gihong Park <https://orcid.org/0009-0003-0632-2684>
 Bokyung Kim <https://orcid.org/0000-0002-9143-9654>
 Hyunsoo Chung <https://orcid.org/0000-0001-5159-357X>
 Sang Gyun Kim <https://orcid.org/0000-0003-1799-9028>
 Soo-Jeong Cho <https://orcid.org/0000-0001-7144-0589>

REFERENCES

1. Reyes VE. *Helicobacter pylori* and its role in gastric cancer. *Microorganisms* 2023;11:1312.
2. Fallone CA, Moss SF, Malfertheiner P. *Helicobacter pylori* infection. *N Engl J Med* 2019;381:587-589.
3. Crowe SE. *Helicobacter pylori* infection. *N Engl J Med* 2019;380:1158-1165.
4. Ford AC, Forman D, Hunt RH, Yuan Y, Moayyedi P. *Helicobacter pylori* eradication therapy to prevent gastric cancer in healthy asymptomatic infected individuals: systematic review and meta-analysis of randomised controlled trials. *BMJ* 2014;348:g3174.
5. Choi IJ, Kook MC, Kim YI, et al. *Helicobacter pylori* therapy for the prevention of metachronous gastric cancer. *N Engl J Med* 2018;378:1085-1095.
6. Jung HK, Kang SJ, Lee YC, et al. Evidence based guidelines for the treatment of *Helicobacter pylori* infection in Korea 2020. *Korean J Intern Med* 2021;36:807-838.
7. Lee JH, Ahn JY, Choi KD, et al. Nationwide antibiotic resistance mapping of *Helicobacter pylori* in Korea: a prospective multicenter study. *Helicobacter* 2019;24:e12592.
8. Malfertheiner P, Megraud F, O'Morain CA, et al. Management of *Helicobacter pylori* infection—the Maastricht V/Florence consensus report. *Gut* 2017;66:6-30.
9. Nishizawa T, Suzuki H. Mechanisms of *Helicobacter pylori* antibiotic resistance and molecular testing. *Front Mol Biosci* 2014;1:19.
10. Srisuphanunt M, Wilairatana P, Kooltheat N, Duangchan T, Katzenmeier G, Rose JB. Molecular mechanisms of antibiotic resistance and novel treatment strategies for *Helicobacter pylori* infections. *Trop Med Infect Dis* 2023;8:163.
11. Lehours P, Siffré E, Mégraud F. DPO multiplex PCR as an alternative to culture and susceptibility testing to detect *Helicobacter pylori* and its resistance to clarithromycin. *BMC Gastroenterol* 2011;11:112.
12. Woo HY, Park DI, Park H, et al. Dual-priming oligonucleotide-based multiplex PCR for the detection of *Helicobacter pylori* and determination of clarithromycin resistance with gastric biopsy specimens. *Helicobacter* 2009;14:22-28.
13. Kim JL, Cho SJ, Chung SJ, et al. Empiric versus clarithromycin resistance-guided therapy for *Helicobacter pylori* based on polymerase chain reaction results in patients with gastric neoplasms or gastric mucosa-associated lymphoid tissue lymphoma: a randomized controlled trial. *Clin Transl Gastroenterol* 2020;11:e00194.
14. Cho SH, Park MS, Park SY, Kim DH, You HS, Kim HS. Effectiveness of 7-day triple therapy with half-dose clarithromycin for the eradication of *Helicobacter pylori* without the A2143G and A2142G point mutations of the 23S rRNA gene in a high clarithromycin resistance area. *Front Med (Lausanne)* 2023;10:1150396.

15. De Francesco V, Margiotta M, Zullo A, et al. Clarithromycin-resistant genotypes and eradication of *Helicobacter pylori*. *Ann Intern Med* 2006;144:94-100.
16. Francavilla R, Lionetti E, Castellaneta S, et al. Clarithromycin-resistant genotypes and eradication of *Helicobacter pylori*. *J Pediatr* 2010; 157:228-232.
17. Kim T, Song HJ, Shin SY, et al. Clarithromycin-resistant *Helicobacter pylori* associated with 23S rRNA point mutations in Jeju Island. *Korean J Gastroenterol* 2013;61:252-258.
18. Kim SY, Park JM, Lim CH, et al. Types of 23S ribosomal RNA point mutations and therapeutic outcomes for *Helicobacter pylori*. *Gut Liver* 2021;15:528-536.
19. Jung MK, Lee JK, Heo J, Kang EJ, Lee YR. The effect of concomitant therapy and quadruple therapy for patients who had 23S ribosomal ribonucleic acid mutated *Helicobacter pylori* in Daegu and Kyungpook Area. *Korean J Helicobacter Up Gastrointest Res* 2014;14:249-254.
20. Lin Y, Shao Y, Yan J, Ye G. Antibiotic resistance in *Helicobacter pylori*: from potential biomolecular mechanisms to clinical practice. *J Clin Lab Anal* 2023;37:e24885.
21. Wang YH, Wang FF, Gong XL, et al. Genotype profiles of *Helicobacter pylori* from gastric biopsies and strains with antimicrobial-induced resistance. *Therap Adv Gastroenterol* 2020;13:1756284820952596.
22. Smith SM, O'Morain C, McNamara D. Antimicrobial susceptibility testing for *Helicobacter pylori* in times of increasing antibiotic resistance. *World J Gastroenterol* 2014;20:9912-9921.
23. Agudo S, Alarcón T, Urruzuno P, Martínez MJ, López-Brea M. Detection of *Helicobacter pylori* and clarithromycin resistance in gastric biopsies of pediatric patients by using a commercially available real-time polymerase chain reaction after NucliSens semiautomated DNA extraction. *Diagn Microbiol Infect Dis* 2010;67:213-219.
24. Binh TT, Shiota S, Suzuki R, et al. Discovery of novel mutations for clarithromycin resistance in *Helicobacter pylori* by using next-generation sequencing. *J Antimicrob Chemother* 2014;69:1796-1803.
25. Hirata K, Suzuki H, Nishizawa T, et al. Contribution of efflux pumps to clarithromycin resistance in *Helicobacter pylori*. *J Gastroenterol Hepatol* 2010;25(Suppl 1):S75-S79.
26. Zhang Z, Liu ZQ, Zheng PY, Tang FA, Yang PC. Influence of efflux pump inhibitors on the multidrug resistance of *Helicobacter pylori*. *World J Gastroenterol* 2010;16:1279-1284.
27. Yonezawa H, Osaki T, Hojo F, Kamiya S. Effect of *Helicobacter pylori* biofilm formation on susceptibility to amoxicillin, metronidazole and clarithromycin. *Microb Pathog* 2019;132:100-108.
28. Salehi N, Attaran B, Zare-Mirakabad F, et al. The outward shift of clarithromycin binding to the ribosome in mutant *Helicobacter pylori* strains. *Helicobacter* 2020;25:e12731.
29. Sakinc T, Baars B, Wüppenhorst N, Kist M, Huebner J, Opferkuch W. Influence of a 23S ribosomal RNA mutation in *Helicobacter pylori* strains on the in vitro synergistic effect of clarithromycin and amoxicillin. *BMC Res Notes* 2012;5:603.
30. Park CG, Kim S, Lee EJ, Jeon HS, Han S. Clinical relevance of point mutations in the 23S rRNA gene in *Helicobacter pylori* eradication: a prospective, observational study. *Medicine (Baltimore)* 2018;97:e11835.
31. Seo SI, Lim H, Bang CS, et al. Bismuth-based quadruple therapy versus metronidazole-intensified triple therapy as a first-line treatment for clarithromycin-resistant *Helicobacter pylori* infection: a multicenter randomized controlled trial. *Gut Liver* 2022;16:697-705.
32. Shrestha AB, Pokharel P, Sapkota UH, et al. Drug resistance patterns of commonly used antibiotics for the treatment of *Helicobacter pylori* infection among South Asian countries: a systematic review and meta-analysis. *Trop Med Infect Dis* 2023;8:172.
33. Ho JJC, Navarro M, Sawyer K, Elfanagely Y, Moss SF. *Helicobacter pylori* antibiotic resistance in the United States between 2011 and 2021: a systematic review and meta-analysis. *Am J Gastroenterol* 2022; 117:1221-1230.